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(54) Title: KETOREDUCTASE GENE AND PROTEIN FROM YEAST

(57) Abstract

This invention provides a cloned ketoreductase gene, vectors for expressing same, recombinant host cells that express said vector-borne gene, and a method for stereospecifically reducing a ketone using a recombinant ketoreductase, or a recombinant host cell that expresses a cloned ketoreductase gene.

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KETOREDUCTASE GENE AND PROTEIN FROM YEAST

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CROSS-REFERENCE

This application claims the benefit of U.S. Provisional Application No. 60/064,195, filed November 4, 1997.

FIELD OF THE INVENTION

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This invention relates to recombinant DNA technology. In particular the invention pertains to the cloning of a ketoreductase gene from Zygosaccharomyces rouxii, and the use of recombinant hosts expressing fungal ketoreductase genes in a process for stereospecific reduction of ketones.

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BACKGROUND OF THE INVENTION

2,3 Benzodiazepine derivatives are potent antagonists of the AMPA (α-amino-3-hydroxy-5 methylisoxazole-4-propionic acid) class of receptors in the mammalian central nervous system (See I. Tarnawa et al. In Amino Acids: Chemistry, Biology and Medicine, Eds. Lubec and Rosenthal, Leiden, 1990). These derivative compounds have potentially widespread applications as neuroprotective agents, particularly as anti-convulsants. One series of 2,3 benzodiazepines is considered particularly advantageous for such use, and this series of compounds has the following general formula:

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Wherein R is hydrogen or C_1 - C_{10} alkyl; and X is hydrogen, C_1 - C_{10} alkyl, acyl, aryl, amido or carboxyl, or a substituted derivative thereof.

The clinical potential for these compounds has led to interest in developing more efficient synthetic methods. Biologically-based methods in which a ketoreductase enzyme provides a stereospecific reduction in a whole-cell process using fungal cells have been described in U.S. Patent application serial number 08/413,036.

BRIEF SUMMARY OF THE INVENTION

The present invention provides isolated nucleic acid molecules that encode a ketoreductase enzyme from Z. rouxii. The invention also provides the protein product of said nucleic acid, in substantially purified form. Also provided are methods for the formation of chiral alcohols using a purified ketoreductase enzyme, or a recombinant host cell that expresses a fungal ketoreductase gene.

Having the cloned ketoreductase gene enables the production of recombinant ketoreductase protein, and the production of recombinant host cells expressing said

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protein, wherein said recombinant cells can be used in a stereospecific reduction of ketones.

In one embodiment the present invention relates to an isolated DNA molecule encoding ketoreductase protein, said DNA molecule comprising the nucleotide sequence identified as SEQ ID NO:1.

In another embodiment the present invention relates to a substantially purified ketoreductase protein molecule from Z. rouxii.

In another embodiment the present invention relates to a ketoreductase protein molecule from Z. rouxii, wherein said protein molecule comprises the sequence identified as SEO ID NO:2.

In a further embodiment the present invention relates
to a ribonucleic acid molecule encoding ketoreductase
protein, said ribonucleic acid molecule comprising the
sequence identified as SEQ ID NO:3.

In yet another embodiment, the present invention relates to a recombinant DNA vector that incorporates a ketoreductase gene in operable-linkage to gene expression sequences, enabling said gene to be transcribed and translated in a host cell.

In still another embodiment the present invention relates to host cells that have been transformed or transfected with a cloned ketoreductase gene such that said ketoreductase gene is expressed in the host cell.

In a still further embodiment, the present invention relates to a method for producing chiral alcohols using recombinant host cells that express an exogenously introduced ketoreductase gene.

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In yet another embodiment, the present invention relates to a method for producing chiral alcohols using recombinant host cells that have been transformed or transfected with a ketoreductase gene from Z. rouxii, or S. cerevisiae.

In yet another embodiment, the present invention relates to a method for producing chiral alcohols using a purified fungal ketoreductase.

DETAILED DESCRIPTION OF THE INVENTION Definitions

SEQ ID NO:1 - SEQ ID NO:3 comprises the DNA, protein, and RNA sequences of ketoreductase from $Z.\ rouxii$.

SEQ ID NO:4- SEQ ID NO:6 comprises the DNA, protein, and RNA sequences of gene YDR541c from S. cerevisiae.

SEQ ID NO:7- SEQ ID NO:9 comprises the DNA, protein, and RNA sequences of YOL151w from S. cerevisiae.

SEQ ID NO:10- SEQ ID NO:12 comprises the DNA, protein, and RNA sequences of YGL157w from S. cerevisiae.

SEQ ID NO:13- SEQ ID NO:15 comprises the DNA, protein, and RNA sequences of YGL039w from S. cerevisiae.

The term "fusion protein" denotes a hybrid protein molecule not found in nature comprising a translational fusion or enzymatic fusion in which two or more different proteins or fragments thereof are covalently linked on a single polypeptide chain.

The term "plasmid" refers to an extrachromosomal genetic element. The starting plasmids herein are either commercially available, publicly available on an

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unrestricted basis, or can be constructed from available plasmids in accordance with published procedures. In addition, equivalent plasmids to those described are known in the art and will be apparent to the ordinarily skilled artisan.

"Recombinant DNA cloning vector" as used herein refers to any autonomously replicating agent, including, but not limited to, plasmids and phages, comprising a DNA molecule to which one or more additional DNA segments can or have been added.

The term "recombinant DNA expression vector" or "expression vector" as used herein refers to any recombinant DNA cloning vector, for example a plasmid or phage, in which a promoter and other regulatory elements are present thereby enabling transcription of an inserted DNA.

The term "vector" as used herein refers to a nucleic acid compound used for introducing exogenous DNA into host cells. A vector comprises a nucleotide sequence which may encode one or more protein molecules. Plasmids, cosmids, viruses, and bacteriophages, in the natural state or which have undergone recombinant engineering, are examples of commonly used vectors.

The terms "complementary" or "complementarity" as used herein refers to the capacity of purine and pyrimidine nucleotides to associate through hydrogen bonding in double stranded nucleic acid molecules. The following base pairs are complementary: guanine and cytosine; adenine and thymine; and adenine and uracil. As used herein "complementary" means that at least one of two hybridizing strands is fully base-paired with the other member of said

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hybridizing strands, and there are no mismatches. Moreover, at each nucleotide position of said one strand, an "A" is paired with a "T", a "T" is paired with an "A", a "G" is paired with a "C", and a "C" is paired with a "G".

"Isolated nucleic acid compound" refers to any RNA or DNA sequence, however constructed or synthesized, which is locationally distinct from its natural location.

A "primer" is a nucleic acid fragment which functions as an initiating substrate for enzymatic or synthetic elongation of, for example, a nucleic acid molecule.

The term "promoter" refers to a DNA sequence which directs transcription of DNA to RNA. An inducible promoter is one that is regulatable by environmental signals, such as carbon source, heat, metal ions, chemical inducers, etc.; a constitutive promoter generally is expressed at a constant level and is not regulatable.

A "probe" as used herein is a labeled nucleic acid compound which can hybridize wih another nucleic acid compound.

The term "hybridization" as used herein refers to a process in which a single-stranded nucleic acid molecule joins with a complementary strand through nucleotide base pairing. "Selective hybridization" refers to hybridization under conditions of high stringency. The degree of hybridization depends upon, for example, the degree of complementarity, the stringency of hybridization, and the length of hybridizing strands.

"Substantially identical" means a sequence having 30 sufficient homology to hybridize under stringent conditions

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and/or be at least 90% identical to a sequence disclosed herein.

The term "stringency" relates to nucleic acid hybridization conditions. High stringency conditions disfavor non-homologous base pairing. Low stringency conditions have the opposite effect. Stringency may be altered, for example, by changes in temperature, denaturants, and salt concentration. Typical high stringency conditions comprise hybridizing at 50°C to 65°C in 5X SSPE and 50% formamide, and washing at 50°C to 65°C in 0.5X SSPE; typical low stringency conditions comprise hybridizing at 35°C to 37° in 5X SSPE and 40% to 45% formamide and washing at 42°C in 1X-2X SSPE.

"SSPE" denotes a hybridization and wash solution

comprising sodium chloride, sodium phosphate, and EDTA, at pH 7.4. A 20% solution of SSPE is made by dissolving 174 g of NaCl, 27.6 g of NaH₂PO4·H₂O, and 7.4 g of EDTA in 800 ml of H₂O. The pH is adjusted with NaOH and the volume brought to 1 liter.

"SSC" denotes a hybridization and wash solution comprising sodium chloride and sodium citrate at pH 7. A 20X solution of SSC is made by dissolving 175 g of NaCl and 88 g of sodium citrate in 800 ml of H₂O. The volume is brought to 1 liter after adjusting the pH with 10N NaOH.

The ketoreductase gene encodes a novel enzyme that catalyzes an asymmetric reduction of selected ketone substrates (See Equation 1 and Table 1).

Equation 1

Table 1: Substrate s	pecificity of	ketoreductase	from Z.	rouxii.
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Compound	Concentration (mM)	% Relative Activity	Compound	Concentration (mM)	% Relative Activity
	3	100	√° 9	3	194
	5	18	0 NH 10	0.8	86
CI OEL	5	42		0.6	17
но о о о о	4	37	12°	5	100
	0.6	4	0 H 13	5	32
	0.6	0	· 7		

The ketoreductase enzymes disclosed herein are members of the carbonyl reductase enzyme class. Carbonyl reductases are involved in the reduction of xenobiotic carbonyl compounds (Hara et. al, Arch. Biochem. Biophys., 244, 238-

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247, 1986) and have been classified into the short-chain dehydrogenase/reductase (SDR) enzyme superfamily (Jörnvall et. al, Biochemistry, 34, 6003-6013, 1995) and the single-domain reductase/epimerase/dehydrogenase (RED) enzyme superfamily (Labesse et. al, Biochem. J., 304, 95-99, 1994). The ketoreductases of this invention are able to effectively reduce a variety of α -ketolactones, α -ketolactams, and diketones (Table 1).

The ketoreductase gene of Z. rouxii comprises a DNA sequence designated herein as SEQ ID NO:1. Those skilled in the art will recognize that owing to the degeneracy of the genetic code (i.e. 64 codons which encode 20 amino acids), numerous "silent" substitutions of nucleotide base pairs could be introduced into the sequence identified as SEQ ID NO:1 without altering the identity of the encoded amino acid(s) or protein product. All such substitutions are intended to be within the scope of the invention.

Gene Isolation Procedures

Those skilled in the art will recognize that the ketoreductase gene may be obtained by a plurality of applicable recombinant DNA techniques including, for example, polymerase chain reaction (PCR) amplification, hybridization to a genomic or cDNA library, or de novo DNA synthesis. (See e.g., J.Sambrook et al. Molecular Cloning, 2d Ed. Chap. 14 (1989)).

Methods for constructing cDNA libraries in a suitable vector such as a plasmid or phage for propagation in procaryotic or eucaryotic cells are well known to those

skilled in the art. [See e.g. J.Sambrook et al. Supra]. Suitable cloning vectors are widely available.

Skilled artisans will recognize that the ketoreductase gene or fragment thereof could be isolated by PCR amplification from a human cDNA library prepared from a 5 tissue in which said gene is expressed, using oligonucleotide primers targeted to any suitable region of SEQ ID NO:1. Methods for PCR amplification are widely known in the art. See e.g. PCR Protocols: A Guide to Method and Application, Ed. M. Innis et.al., Academic Press (1990). The 10 amplification reaction comprises template DNA, suitable enzymes, primers, nucleoside triphosphates, and buffers, and is conveniently carried out in a DNA Thermal Cycler (Perkin Elmer Cetus, Norwalk, CT). A positive result is determined by detecting an appropriately-sized DNA fragment following 15 gel electrophoresis.

Protein Production Methods

One embodiment of the present invention relates to
the substantially purified ketoreductase enzyme (identified herein as SEQ ID NO:2) encoded by the Z. rouxii
ketoreductase gene (identified herein as SEQ ID NO:1).

Skilled artisans will recognize that the proteins of the present invention can be synthesized by a number of different methods, such as chemical methods well known in the art, including solid phase peptide synthesis or recombinant methods. Both methods are described in U.S. Patent 4,617,149, incorporated herein by reference. The proteins of the invention can also be purified by well known

methods from a culture of cells that produce the protein, for example, Z. rouxii.

The principles of solid phase chemical synthesis of polypeptides are well known in the art and may be found in general texts in the area. See, e.g., H. Dugas and C. Penney, <u>Bioorganic Chemistry</u> (1981) Springer-Verlag, New York, 54-92. For example, peptides may be synthesized by solid-phase methodology utilizing an Applied Biosystems 430A peptide synthesizer (Applied Biosystems, Foster City, CA) and synthesis cycles supplied by Applied Biosystems.

The protein of the present invention can also be produced by recombinant DNA methods using the cloned ketoreductase gene. Recombinant methods are preferred if a high yield is desired. Expression of the cloned gene can be carried out in a variety of suitable host cells, well known to those skilled in the art. For this purpose, the ketoreductase gene is introduced into a host cell by any suitable means, well known to those skilled in the art. While chromosomal integration of the cloned gene is within the scope of the present invention, it is preferred that the gene be cloned into a suitable extra-chromosomally maintained expression vector so that the coding region of the ketoreductase gene is operably-linked to a constitutive or inducible promoter.

The basic steps in the recombinant production of the ketoreductase protein are:

a) constructing a natural, synthetic or semi-synthetic DNA encoding ketoreductase protein;

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- b) integrating said DNA into an expression vector in a manner suitable for expressing the ketoreductase protein, either alone or as a fusion protein; or integrating said DNA into a host chromosome such that said DNA expresses ketoreductase;
- c) transforming or otherwise introducing said vector into an appropriate eucaryotic or prokaryotic host cell forming a recombinant host cell,
- d) culturing said recombinant host cell in a manner to express the ketoreductase protein; and
- e) recovering and substantially purifying the ketoreductase protein by any suitable means, well known to those skilled in the art.

Expressing Recombinant ketoreductase Protein in Procaryotic and Eucaryotic Host Cells

procaryotes may be employed in the production of
the ketoreductase protein. For example, the Escherichia
coli K12 strain 294 (ATCC No. 31446) or strain RV308 is
particularly useful for the prokaryotic expression of
foreign proteins. Other strains of E. coli, bacilli such as
Bacillus subtilis, enterobacteriaceae such as Salmonella
typhimurium or Serratia marcescans, various Pseudomonas

species and other bacteria, such as *Streptomyces*, may also be employed as host cells in the cloning and expression of the recombinant proteins of this invention.

Promoter sequences suitable for driving the expression of genes in procaryotes include $\;\beta$ -lactamase 5 [e.g. vector pGX2907, ATCC 39344, contains a replicon and β -lactamase gene], lactose systems [Chang et al., Nature (London), 275:615 (1978); Goeddel et al., Nature (London), 281:544 (1979)], alkaline phosphatase, and the tryptophan (trp) promoter system [vector pATH1 (ATCC 37695) which is 10 designed to facilitate expression of an open reading frame as a trpE fusion protein under the control of the trp promoter]. Hybrid promoters such as the tac promoter (isolatable from plasmid pDR540, ATCC-37282) are also suitable. Still other bacterial promoters, whose nucleotide 15 sequences are generally known, enable one of skill in the art to ligate such promoter sequences to DNA encoding the proteins of the instant invention using linkers or adapters to supply any required restriction sites. Promoters for use in bacterial systems also will contain a Shine-Dalgarno 20 sequence operably linked to the DNA encoding the desired polypeptides. These examples are illustrative rather than limiting.

The protein(s) of this invention may be

synthesized either by direct expression or as a fusion
protein comprising the protein of interest as a
translational fusion with another protein or peptide which
may be removable by enzymatic or chemical cleavage. It is
often observed in the production of certain peptides in
recombinant systems that expression as a fusion protein

prolongs the lifespan, increases the yield of the desired peptide, or provides a convenient means of purifying the protein. A variety of peptidases (e.g. enterokinase and thrombin) which cleave a polypeptide at specific sites or digest the peptides from the amino or carboxy termini (e.g. diaminopeptidase) of the peptide chain are known. Furthermore, particular chemicals (e.g. cyanogen bromide) will cleave a polypeptide chain at specific sites. skilled artisan will appreciate the modifications necessary to the amino acid sequence (and synthetic or semi-synthetic 10 coding sequence if recombinant means are employed) to incorporate site-specific internal cleavage sites. See e.g., P. Carter, "Site Specific Proteolysis of Fusion Proteins", Chapter 13, in Protein Purification: From Molecular Mechanisms to Large Scale Processes, American 15 Chemical Society, Washington, D.C. (1990).

In addition to procaryotes, a variety of
eucaryotic microorganisms including yeast are suitable host
cells. The yeast Saccharomyces cerevisiae is the most

20 commonly used eucaryotic microorganism. Other yeasts such as
Kluyveromyces lactis, Schizosaccharomyces pombe, and Pichia
pastoris are also suitable. For expression in
Saccharomyces, the plasmid YRp7 (ATCC-40053), for example,
may be used. See, e.g., L. Stinchcomb, et al., Nature,
25 282:39 (1979); J. Kingsman et al., Gene, 7:141 (1979); S.
Tschemper et.al., Gene, 10:157 (1980). Plasmid YRp7
contains the TRP1 gene which provides a selectable marker
for use in a trpl auxotrophic mutant.

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An expression vector carrying a cloned ketoreductase gene is transformed or transfected into a suitable host cell using standard methods. Host cells may comprise procaryotes, such as E. coli, or simple eucaryotes, such as Z. rouxii, S. cerevisiae, S. pombe, P. pastoris, and K. Lactis. Cells which contain the vector are propagated under conditions suitable for expression of an encoded ketoreductase protein. If the recombinant gene has been placed under the control of an inducible promoter then suitable growth conditions would incorporate the appropriate inducer. The recombinantly-produced protein may be purified from cellular extracts of transformed cells by any suitable means.

In a preferred process for protein purification,

the ketoreductase gene is modified at the 5' end to
incorporate several histidine residues at the amino terminus
of the ketoreductase protein product. This "histidine tag"
enables a single-step protein purification method referred
to as "immobilized metal ion affinity chromatography"

(IMAC), essentially as described in U.S. Patent 4,569,794
which hereby is incorporated by reference. The IMAC method
enables rapid isolation of substantially pure ketoreductase
protein starting from a crude cellular extract.

Other embodiments of the present invention

25 comprise isolated nucleic acid sequences which encode SEQ ID NO:2. As skilled artisans will recognize, the amino acid compounds of the invention can be encoded by a multitude of different nucleic acid sequences because most of the amino acids are encoded by more than one codon. Because these

30 alternative nucleic acid sequences would encode the same

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amino acid sequences, the present invention further comprises these alternate nucleic acid sequences.

The ketoreductase genes disclosed herein, for example SEQ ID NO:1, may be produced using synthetic methodology. The synthesis of nucleic acids is well known in the art. See, e.g., E.L. Brown, R. Belagaje, M.J. Ryan, and H.G. Khorana, Methods in Enzymology, 68:109-151 (1979). A DNA segment corresponding to a ketoreductase gene could be generated using a conventional DNA synthesizing apparatus, such as the Applied Biosystems Model 380A or 380B DNA synthesizers (Applied Biosystems, Inc., 850 Lincoln Center Drive, Foster City, CA 94404) which employ phosphoramidite chemistry. Alternatively, phosphotriester chemistry may be employed to synthesize the nucleic acids of this invention. [See, e.g., M.J. Gait, ed., Oligonucleotide Synthesis, A

Practical Approach, (1984).]

In an alternative methodology, namely PCR, a DNA

sequence comprising a portion or all of SEQ ID NO:1, SEQ ID NO:4, SEQ ID NO:7, SEQ ID NO:10, or SEQ ID NO:13 can be generated from a suitable DNA source, for example Z. rouxii or S. cerevisiae genomic DNA or cDNA. For this purpose, suitable oligonucleotide primers targeting SEQ ID NO:1, SEQ ID NO:4, SEQ ID NO:7, SEQ ID NO:10, SEQ ID NO:13 or region therein are prepared, as described in U.S. Patent No.

4,889,818, which hereby is incorporated by reference.

Protocols for performing the PCR are disclosed in, for example, PCR Protocols: A Guide to Method and Applications, Ed. Michael A. Innis et al., Academic Press, Inc. (1990).

The ribonucleic acids of the present invention may

30 be prepared using the polynucleotide synthetic methods

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discussed *supra*, or they may be prepared enzymatically using RNA polymerase to transcribe a ketoreductase DNA template.

See e.g., J. Sambrook, et. al., supra, at 18.82-18.84.

This invention also provides nucleic acids, RNA or DNA, which are complementary to SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:13, or SEQ ID NO:15.

The present invention also provides probes and primers useful for a variety of molecular biology techniques including, for example, hybridization screens of genomic, subgenomic, or cDNA libraries. A nucleic acid compound comprising SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:13, or SEQ ID NO:15, or a complementary sequence thereof, or a fragment thereof, which is at least 18 base pairs in length, and which will selectively hybridize to DNA encoding a ketoreductase, is provided. Preferably, the 18 or more base pair compound is DNA. See e.g. B. Wallace and G. Miyada, "Oligonucleotide Probes for the Screening of Recombinant DNA Libraries," In Methods in Enzymology, Vol. 152, 432-442, Academic Press (1987).

Probes and primers can be prepared by enzymatic methods well known to those skilled in the art (See e.g. Sambrook et al. supra). In a most preferred embodiment these probes and primers are synthesized using chemical means as described above.

Another aspect of the present invention relates to recombinant DNA cloning vectors and expression vectors comprising the nucleic acids of the present invention. The preferred nucleic acid vectors are those which comprise DNA.

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The most preferred recombinant DNA vectors comprise a isolated DNA sequence selected from the group consisting of SEQ ID NO:1, SEQ ID NO:4, SEQ ID NO:7, SEQ ID NO:10, or SEQ ID NO:13.

The skilled artisan understands that choosing the most appropriate cloning vector or expression vector depends upon a number of factors including the availability of restriction enzyme sites, the type of host cell into which the vector is to be transfected or transformed, the purpose of the transfection or transformation (e.g., stable transformation as an extrachromosomal element, or integration into the host chromosome), the presence or absence of readily assayable or selectable markers (e.g., antibiotic resistance and metabolic markers of one type and another), and the number of copies of the gene to be present in the host cell.

Vectors suitable to carry the nucleic acids of the present invention comprise RNA viruses, DNA viruses, lytic bacteriophages, lysogenic bacteriophages, stable bacteriophages, plasmids, viroids, and the like. The most preferred vectors are plasmids.

When preparing an expression vector the skilled artisan understands that there are many variables to be considered, for example, whether to use a constitutive or inducible promoter. Inducible promoters are preferred because they enable high level, regulatable expression of an operably-linked gene. Constitutive promoters are further suitable in instances for which secretion or extra-cellular export is desireable. The skilled artisan will recognize a number of inducible promoters which respond to a variety of

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inducers, for example, carbon source, metal ions, and heat. The practitioner also understands that the amount of nucleic acid or protein to be produced dictates, in part, the selection of the expression system. The addition of certain nucleotide sequences is useful for directing the localization of a recombinant protein. For example, a sequence encoding a signal peptide preceding the coding region of a gene, is useful for directing the extra-cellular export of a resulting polypeptide.

Host cells harboring the nucleic acids disclosed herein are also provided by the present invention. Suitable host cells include procaryotes, such as *E. coli*, or simple eucaryotes, such as fungal cells, which have been transfected or transformed with a vector which comprises a nucleic acid of the present invention.

The present invention also provides a method for constructing a recombinant host cell capable of expressing SEQ ID NO:2, SEQ ID NO:5, SEQ ID NO:8, SEQ ID NO:11, or SEQ ID NO:14, said method comprising transforming or otherwise introducing into a host cell a recombinant DNA vector that comprises an isolated DNA sequence which encodes SEQ ID NO:2, SEQ ID NO:5, SEQ ID NO:8, SEQ ID NO:11, or SEQ ID NO:14. Preferred vectors for expression are those which comprise SEQ ID NO:1. Transformed host cells may be cultured under conditions well known to skilled artisans such that SEQ ID NO:2, SEQ ID NO:5, SEQ ID NO:8, SEQ ID NO:11, or SEQ ID NO:14 is expressed, thereby producing a ketoreductase protein in the recombinant host cell.

For the purpose of identifying or developing inhibitors or other modifiers of the enzymes disclosed

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herein, or for identifying suitable substrates for bioconversion, it would be desirable to identify compounds that bind and/or inhibit, or otherwise modify, the ketoreductase enzyme and its associated activity. A method for determining agents that will modify the ketoreductase activity comprises contacting the ketoreductase protein with a test compound and monitoring the alteration of enzyme activity by any suitable means.

The instant invention provides such a screening
system useful for discovering compounds which bind the
ketoreductase protein, said screening system comprising the
steps of:

- a) preparing ketoreductase protein;
- b) exposing said ketoreductase protein to a test compound;
- c) quantifying a modulation of activity by said compound.

Utilization of the screening system described above provides a means to determine compounds which may alter the activity of ketoreductase. This screening method may be adapted to automated procedures such as a PANDEX® (Baxter-Dade Diagnostics) system, allowing for efficient high-volume screening of potential modifying agents.

In such a screening protocol, ketoreductase is
prepared as described herein, preferably using recombinant
DNA technology. A test compound is introduced into a
reaction vessel containing ketoreductase, followed by
addition of enzyme substrate. For convenience the reaction
can be coupled to the oxidation of NADPH, thereby enabling

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progress to be monitored spectrophotometrically by measuring the absorbance at 340 nm. Alternatively, substrate may be added simultaneously with a test compound. In one method radioactively or chemically-labeled compound may be used. The products of the enzymatic reaction are assayed for the chemical label or radioactivity by any suitable means. The absence or diminution of the chemical label or radioactivity indicates the degree to which the reaction is inhibited.

The following examples more fully describe the present invention. Those skilled in the art will recognize that the particular reagents, equipment, and procedures described are merely illustrative and are not intended to limit the present invention in any manner.

15 EXAMPLE 1

Construction of a DNA Vector for Expressing a Ketoreductase Gene in a Homologous or Heterologous Host

A plasmid comprising the Z. rouxii ketoreductase gene suitable for expressing said gene in a host cell, for example E. coli (DE3) strains, contains an origin of replication (Ori), an ampicillin resistance gene (Amp), useful for selecting cells which have incorporated the vector following a tranformation procedure, and further comprises the lacI gene for repression of the lac operon, as well as the T7 promoter and T7 terminator sequences in operable linkage to the coding region of the ketoreductase gene. Parent plasmid pET11A (obtained from Novogen, Madison, WI) was linearized by digestion with endonucleases NdeI and BamHI. Linearized pET11A was ligated to a DNA fragment

bearing NdeI and BamHI sticky ends and further comprising the coding region of the Z.rouxii ketoreductase gene.

The ketoreductase gene is isolated most conveniently by the PCR. Genomic DNA from Z. rouxii isolated by standard methods was used for amplification of the ketoreductase gene. Primers are synthesized corresponding to the 5' and 3' ends of the gene (SEQ ID NO:1) to enable amplification of the coding region.

The ketoreductase gene (nucleotides 164 through 1177 of SEQ ID NO:1) ligated into the vector was modified at the 5' end (amino terminus of encoded protein) in order to simplify purification of the encoded ketoreductase protein. For this purpose, an oligonucleotide encoding 8 histidine residues and a factor Xa cleavage site was inserted after the ATG start codon at nucleotide positions 164 to 166 of SEQ ID NO:1. Placement of the histidine residues at the amino terminus of the encoded protein does not affect its activity and serves only to enable the IMAC one-step protein purification procedure.

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EXAMPLE 2

Purification of Ketoreductase from Z. rouxii

Approximately 1 gram of Z. rouxii cell paste was resuspended in Lysing Buffer, comprising 50 mM Tris-Cl pH 7.5, 2 mM EDTA supplemented with pepstatin (1 μ g/mL), leupeptin (1.25 μ g/mL), aprotinin (2.5 μ g/mL), and AEBSF (25 μ g/mL). The cells were lysed using a DynoMill (GlenMills, Inc. Clifton, NJ) equipped with 0.5-0.75 mm lead free beads under continuous flow conditions according to the manufacturer's recommended use. After four complete passes

through the DynoMill, the material was centrifuged twice (25,000 x g for 30 minutes at 4°C). Solid ammonium sulfate (291 g/liter) was added slowly to the resulting clarified cell extract with stirring at 4°C to achieve 50% saturation. After 1 hour, the mixture was centrifuged at 23,000 x g for 5 The supernatant was then brought to 85% saturation by the addition of solid ammonium sulfate (159 q/liter) and stirred for 1h at 4°C before centrifugation (23,000 xg for 30 min). The resultant 50-85% ammonium sulfate pellet was resuspended in 600 mL of Lysing Buffer 10 and the residual ammonium sulfate was removed by dialysis against the same buffer at 4°C. The desalted material was centrifuged twice to remove particulate matter (23,000 xq for 30 min) and 700 - 800 Units of the clarified material was loaded onto a Red-120 dye affinity column (32 mm X 140 15 mm) equilibrated in 50 mM Tris-Cl pH 7.5, 1 mM MgCl₂, pepstatin (1 μ g/mL), leupeptin (1.25 μ g/mL), and aprotinin (2.5 μ g/mL). Reductase activity was eluted from the column at a flowrate of 8 mL/min under the following conditions: 1) a 10 minute linear gradient from 0 - 0.3 M NaCl; 2) 13 20 minutes at 0.3 M NaCl; 3) a 60 minute linear gradient from 0.3 - 1.5 M NaCl. The fractions containing reductase activity were pooled, and changed to 20 mM potassium phosphate buffer (pH 7.2), pepstatin (1 μ g/mL), leupeptin (1.25 μ g/mL), and aprotinin (2.5 μ g/mL) by dialysis at 4°C. 25 The sample was clarified by centrifugation (23,000 \times g for 30 min) and 400 Units was loaded onto a Bio-Scale CHT-I hydroxyapatite column (15 mm x 113 mm, Bio-Rad, Inc.) equilibrated in the same buffer that had been made 5% in glycerol. Reductase activity was eluted from the column at 30

a flowrate of 5.0 mL/min in a sodium chloride step gradient consisting of 5 minutes at 0 M NaCl, a gradient step to 0.7 M NaCl which was maintained for 10 minutes, and then a 20 minute linear gradient from 0.7 - 1.0 M NaCl. The fractions containing reductase activity were pooled and desalted with 20 mM potassium phosphate buffer (pH 7.2), pepstatin A (1 μ g/mL), leupeptin (1.25 μ g/mL), and aprotinin (2.5 μ g/mL) by dialysis at 4°C. The sample (100- 200 Units) was loaded onto a Bio-Scale CHT-I hydroxyapatite column (10 mm x 64 mm) equilibrated in the same buffer which had been made 5% in Reductase activity was eluted from the column at a flowrate of 2.0 mL/min in a 25 minute linear gradient from 0 to 50% 400 mM potassium phosphate (pH 6.8), 5% glycerol. Fractions containing reductase activity were pooled and changed into 10 mM Tris-Cl (pH 8.5) by dialysis at 4°C. sample was then made 10% in glycerol, concentrated to 0.4 mg/mL by ultrafiltration (Amicon, YM-10), and stored at -70°C.

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EXAMPLE 3

Reductase Activity Using the Ketoreductase from Z. rouxii

Reductase activity was measured using a suitable substrate and a partially purified or substantially purified ketoreductase from Z. rouxii. Activity was measured as a function of the absorbance change at 340 nm, resulting from the oxidation of NADPH. The 1 ml assay contained a mixture of 3.0 mM 3,4-methylenedioxyphenyl acetone, 162 μ M NADPH, 50 mM MOPS buffer (pH 6.8), and 0.6 mU of ketoreductase and was carried out at 26° C. Reaction mixtures were first equilibrated at 26°C for 10 min in the absence of NADPH, and

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then initiated by addition of NADPH. The absorbance was measured at 340 nm every 15 seconds over a 5 minute period; the change in absorbance was found to be linear over that time period. The kinetic parameters for 3,4methylenedioxyphenyl acetone were determined at an NADPH concentration of 112 μM and a 3,4-methylenedioxyphenyl acetone concentration that varied from 1.7 mM - 7.2 mM. The kinetic parameters for NADPH were determined by maintaining the 3,4-methylenedioxyphenyl acetone concentration at 3 mM and the NADPH concentration was varied from 20.5 μM - 236.0 An extinction coefficient of 6220 M⁻¹ cm⁻¹ for NADPH absorbance at 340 nm was used to calculate the specific activity of the enzyme. For assays using isatin, the change in absorbance with time was measured at 414 nm using an extinction coefficient of 849 ${\rm M}^{-1}~{\rm cm}^{-1}$ to calculate activity. One Unit of activity corresponds to 1 μ mol of NADPH consumed per minute. For assays carried out at differing pH values, 10 mM Bis-Tris and 10 mM Tris were adjusted to the appropriate pH with HCl. Kinetic parameters were determined by non-linear regression using the JMP® statistics and graphics program.

EXAMPLE 4

Whole Cell Method for Stereoselective Reduction of Ketone Using Recombinant Yeast Cell

A vector for expressing the cloned Z. rouxii ketoreductase gene (SEQ ID NO:1) in a procaryotic or fungal cell, such as S. cerevisiae, is constructed as follows. A 1014 base pair fragment of Z rouxii genomic DNA or cDNA, carrying the ketoreductase gene, is amplified by PCR using

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primers targeted to the ends of the coding region specified in SEQ ID NO:1. It is desireable that the primers also incorporate suitable cloning sites for cloning of said 1014 base pair fragment into an expression vector. The appropriate fragment encoding ketoreductase is amplified and purified using standard methods, for cloning into an expression vector.

A suitable vector for expression in *E. coli* and *S. cerevisiae* is pYX213 (available from Novagen, Inc., 597
Science Drive, Madison, WI 53711; Code MBV-029-10), a 7.5
Kb plasmid that carries the following genetic markers: ori, 2μ circle, Amp^R, CEN, URA3, and the GAL promoter, for high level expression in yeast. Downstream of the GAL promoter, pYX213 carries a multiple cloning site (MCS), which will accommodate the ketoreductase gene amplified in the preceding step. A recombinant plasmid is created by digesting pYX213 and the amplified ketoreductase gene with a restriction enzyme, such as BamH1, and ligating the fragments together.

A recombinant expression vector carrying the Z.rouxii ketoreductase gene is transformed into a suitable Ura strain of S. cerevisiae, using well known methods. Ura transformants are selected on minimal medium lacking uracil.

Expression of the recombinant ketoreductase gene

25 may be induced if desired by growing transformants in

minimal medium that contains 2% galactose as the sole carbon

source.

To carry out a whole cell stereospecific reduction, 3,4-methylenedioxyphenyl acetone is added to a culture of transformants to a concentration of about 10

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grams per liter of culture. The culture is incubated with shaking at room temperature for 24 hours, and the presence of the chiral alcohol analyzed by HPLC.

WE CLAIM:

- I. A substantially pure ketoreductase protein having the amino acid sequence which is SEQ ID NO:2.
- 2. An isolated nucleic acid compound encoding the protein of Claim 1, said protein having the amino acid sequence which is SEQ ID NO:2.
- 3. An isolated nucleic acid compound encoding the protein of Claim 1, wherein said compound has a sequence selected from the group consisting of:
 - (a) SEQ ID NO:1; or
 - (b) SEQ ID NO:3.
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- 4. An isolated nucleic acid compound of Claim 3 wherein the sequence of said compound is SEQ ID NO:1
- 5. An isolated nucleic acid compound having a sequence complementary to SEQ ID NO:1.
 - 6. An isolated nucleic acid compound of Claim 3 wherein the sequence of said compound is SEQ ID NO:3.
- 7. An isolated nucleic acid compound having a sequence complementary to SEQ ID NO:3.
 - 8. A vector comprising an isolated nucleic acid compound of Claim 2.
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- 9. A vector comprising an isolated nucleic acid compound of Claim 3.
- 10. A vector of Claim 9, wherein said isolated nucleic acid compound is SEQ ID NO:1 operably-linked to a promoter sequence.

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- 11. A host cell containing the vector of Claim 10.
- 12. A method for constructing a recombinant host cell having the potential to express SEQ ID NO:2, said method comprising introducing into said host cell by any suitable means a vector of Claim 9.
- 13. A method for expressing SEQ ID NO:2 in the recombinant
 10 host cell of Claim 12, said method comprising culturing said
 recombinant host cell under conditions suitable for gene
 expression.
- 14. A method for reducing a ketone in a stereospecific

 15 manner comprising providing a quantity of a suitable ketone
 to a culture of recombinant cells for a suitable period of
 time, wherein said cells are transformed with a vector that
 carries a ketoreductase gene, and wherein said cells express
 said ketoreductase gene.
 - 15. A method, as in claim 14 wherein said gene is selected from the group consisting of SEQ ID NO:1, SEQ ID NO:4, SEQ ID NO:7, SEQ ID NO:10, and SEQ ID NO:13.
- 25 16. A method, as in claim 14 wherein said ketone comprises an α -ketolactone, α -ketolactam, or a diketone.
 - 17. A method, as in Claim 14, wherein said recombinant cells are selected from the group consisting of *S. cerevisiae*, *Z. rouxii*, and *E. coli*.
 - 18. A method for reducing a ketone in a stereospecific manner comprising mixing a quantity of a suitable ketone with a substantially purified ketoreductase and suitable reducing agent.

- 19. A method, as in Claim 18 wherein said ketoreductase is selected from the group consisting of SEQ ID NO:2, SEQ ID NO:5, SEQ ID NO:11, and SEQ ID NO:14.
- 20. An isolated nucleic acid compound that encodes a protein having ketoreductase activity wherein said nucleic acid hybridizes under high stringency conditions to SEQ ID NO:1, SEQ ID NO:4, SEQ ID NO:7, SEQ ID NO:10, or SEQ ID NO:13.
- 10 21. A method, as in Claim 18 wherein said reducing agent is NADPH.

- 1 -

SEQUENCE LISTING

- (1) GENERAL INFORMATION:
 - (i) APPLICANT: Costello, Colleen A. Menke, Michael A. Hershberger, Charles L. Zmijewski, Milton J.
 - (ii) TITLE OF INVENTION: Ketoreductase Gene and Protein From Yeast
 - (iii) NUMBER OF SEQUENCES: 15
 - (iv) CORRESPONDENCE ADDRESS:
 - (A) ADDRESSEE: Eli Lilly and Company
 - (B) STREET: Lilly Corporate Center
 - (C) CITY: Indianapolis
 - (D) STATE: Indiana
 - (E) COUNTRY: United States
 - (F) ZIP: 46285
 - (v) COMPUTER READABLE FORM:
 - (A) MEDIUM TYPE: Floppy disk
 - (B) COMPUTER: IBM PC compatible
 - (C) OPERATING SYSTEM: PC-DOS/MS-DOS
 - (D) SOFTWARE: PatentIn Release #1.0, Version #1.30
 - (vi) CURRENT APPLICATION DATA:
 - (A) APPLICATION NUMBER:
 - (B) FILING DATE:
 - (C) CLASSIFICATION:
 - (viii) ATTORNEY/AGENT INFORMATION:
 - (A) NAME: Webster, Thomas D.
 - (B) REGISTRATION NUMBER: 39,872
 - (C) REFERENCE/DOCKET NUMBER: X-11325
 - (ix) TELECOMMUNICATION INFORMATION:
 - (A) TELEPHONE: 317-276-3334
 - (2) INFORMATION FOR SEQ ID NO:1:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1270 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: DNA (genomic)
 - (iii) HYPOTHETICAL: NO
 - (iv) ANTI-SENSE: NO

- 2 -

(IX) PEATURE:	lί	x)	FEATURE	:
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(A) NAME/KEY: CDS

(B) LOCATION: 164..1177

(D) OTHER INFORMATION: Z.rouxii ketoreductase

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

	(XI)	SEÇ	OENC	בע ב	SCKI	FIIO	D	- X	2 2 1 0	•							
TGAA'	TGGT	TA I	TTTA	GCAA	T TG	CTGT	GTGA	. GGC	ACTG	ACC	TAAA	GATG	TG I	'ATA	ATAGT	ı	60
GGGA	CTGT	GT A	CTCA	TGAG	G AT	CAAT	ACAT	GTA	AAAT	CTT	ACCA	TACI	TT C	ACAC	AAGTC	!	120
AACT	TAGA	AT (CAATC	AATC	A AT	'CAAT	ТААТ	CAA	GCTA	TAC	AAT	ATG Met 1	ACA Thr	AAA Lys	GTC Val		175
TTC Phe 5	GTA Val	ACA Thr	GGT Gly	GCC Ala	AAC Asn 10	GGA Gly	TTC Phe	GTT Val	GCT Ala	CAA Gln 15	CAC His	GTC Val	GTT Val	CAT His	CAA Gln 20		223
CTA Leu	TTA Leu	GAA Glu	AAG Lys	AAC Asn 25	TAT Tyr	ACA Thr	GTG Val	GTT Val	GGA Gly 30	TCT Ser	GTC Val	CGT Arg	TCA Ser	ACT Thr 35	GAG Glu		271
AAA Lys	GGT Gly	GAT Asp	AAA Lys 40	TTA Leu	GCT Ala	AAA Lys	TTG Leu	CTA Leu 45	AAC Asn	AAT Asn	CCA Pro	AAA Lys	TTT Phe 50	TCA Ser	TAT Tyr		319
GAG Glu	ATT Ile	ATT Ile 55	AAA Lys	GAT Asp	ATG Met	GTC Val	AAT Asn 60	TCG Ser	AGA Arg	GAT Asp	GAA Glu	TTC Phe 65	GAT Asp	AAG Lys	GCT Ala		367
TTA Leu	CAA Gln 70	AAA Lys	CAT His	TCA Ser	GAT Asp	GTT Val 75	GAA Glu	ATT Ile	GTC Val	TTA Leu	CAT His 80	ACT Thr	GCT Ala	TCA Ser	CCA Pro		415
GTC Val 85	TTC Phe	CCA Pro	GGT Gly	GGT Gly	ATT Ile 90	AAA Lys	GAT Asp	GTT Val	GAA Glu	AAA Lys 95	GAA Glu	ATG Met	ATC Ile	CAA Gln	CCA Pro 100		463
GCT Ala	GTT Val	AAT Asn	GGT Gly	ACT Thr 105	Arg	AAT Asn	GTC Val	TTG Leu	TTA Leu 110	TCA Ser	ATC Ile	AAG Lys	GAT Asp	AAC Asn 115	TTA Leu		511
CCA Pro	AAT Asn	GTC Val	AAG Lys 120	Arg	Phe	Val	Tyr	Thr	Ser	Ser	TTA Leu	Ala	GCT Ala 130	Val	CGT Arg		559
ACT Thr	GAA Glu	GGT Gly 135	r GCT / Ala	GGT Gly	TAT	AGT Ser	GCA Ala 140	Asp	GAA Glu	GTI Val	GTC Val	ACC Thr 145	Glu	GAT Asp	TCT Ser		607
TGG Trp	AAC Asr	AA!	r ATT	GCF	TTG Leu	AAA Lys	GAT Asp	GCC Ala	ACC Thr	AAG Lys	GAT Asp	GAA Glu	GGT	ACA Thr	GCT Ala		655

- 3 -

	150					155					160					
TAT Tyr 165	GAG Glu	GCT Ala	TCC Ser	AAG Lys	ACA Thr 170	TAT Tyr	GGT Gly	GAA Glu	AAA Lys	GAA Glu 175	GTT Val	TGG Trp	AAT Asn	TTC Phe	TTC Phe 180	703
GAA Glu	AAA Lys	ACT Thr	AAA Lys	AAT Asn 185	GTT Val	AAT Asn	TTC Phe	GAT Asp	TTT Phe 190	GCC Ala	ATC Ile	ATC Ile	AAC Asn	CCA Pro 195	GTT Val	751
TAT Tyr	GTC Val	TTT Phe	GGT Gly 200	CCT Pro	CAA Gln	TTA Leu	TTT Phe	GAA Glu 205	GAA Glu	TAC Tyr	GTT Val	ACT Thr	GAT Asp 210	AAA Lys	TTG Leu	799
AAC Asn	TTT Phe	TCC Ser 215	AGT Ser	GAA Glu	ATC Ile	ATT Ile	AAT Asn 220	AGT Ser	ATA Ile	ATA Ile	AAA Lys	GGT Gly 225	GAA Glu	AAG Lys	AAG Lys	847
GAA Glu	ATT Ile 230	GAA Glu	GGT Gly	TAT Tyr	GAA Glu	ATT Ile 235	GAT Asp	GTT Val	AGA Arg	GAT Asp	ATT Ile 240	GCA Ala	AGA Arg	GCT Ala	CAT His	895
ATC Ile 245	TCT Ser	GCT Ala	GTT Val	GAA Glu	AAT Asn 250	CCA Pro	GCA Ala	ACT Thr	ACA Thr	CGT Arg 255	CAA Gln	AGA Arg	TTA Leu	ATT Ile	CCA Pro 260	943
GCA Ala	GTT Val	GCA Ala	CCA Pro	TAC Tyr 265	AAT Asn	CAA Gln	CAA Gln	ACT Thr	ATC Ile 270	TTG Leu	GAT Asp	GTT Val	TTG Leu	AAT Asn 275	GAA Glu	991
AAC Asn	TTC Phe	CCA Pro	GAA Glu 280	TTG Leu	AAA Lys	GGT Gly	AAA Lys	ATC Ile 285	GAT Asp	GTT Val	GGG	AAA Lys	CCA Pro 290	GGT Gly	TCT Ser	1039
CAA Gln	AAT Asn	GAA Glu 295	Phe	ATT Ile	AAA Lys	AAA Lys	TAT Tyr 300	TAT Tyr	AAA Lys	TTA Leu	GAT Asp	AAC Asn 305	TCA Ser	AAG Lys	ACC Thr	1087
AAA Lys	AAA Lys 310	Val	TTA Leu	GGT Gly	TTT Phe	GAA Glu 315	Phe	ATT Ile	TCC Ser	CAA Gln	GAG Glu 320	Gln	ACA Thr	ATC Ile	AAA Lys	1135
GAT Asp 325	Ala	GCT Ala	GCT	CAA Gln	ATC Ile 330	Leu	TCC Ser	GTT Val	AAA Lys	AAT Asn 335	Gly	AAA Lys	AAA Lys			1177
TAA	GTGA	ACT	AGAC	CTGI	CA C	TATC	AGAT	T AT	TAGA	GTTC	TGT	ATAG	ATT	AAAG'	IGTGAA	1237
LAA	GTAT	TAG	AATO	ATA	TT I	TATA	ATAT	rg cc	T							1270

(2) INFORMATION FOR SEQ ID NO:2:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 338 amino acids

- 4 -

- (B) TYPE: amino acid
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:
- Met Thr Lys Val Phe Val Thr Gly Ala Asn Gly Phe Val Ala Gln His
- Val Val His Gln Leu Leu Glu Lys Asn Tyr Thr Val Val Gly Ser Val 20 25 30
- Arg Ser Thr Glu Lys Gly Asp Lys Leu Ala Lys Leu Leu Asn Asn Pro
- Lys Phe Ser Tyr Glu Ile Ile Lys Asp Met Val Asn Ser Arg Asp Glu
 50 55 60
- Phe Asp Lys Ala Leu Gln Lys His Ser Asp Val Glu Ile Val Leu His 65 70 75 80
- Thr Ala Ser Pro Val Phe Pro Gly Gly Ile Lys Asp Val Glu Lys Glu 85 90 95
- Met Ile Gln Pro Ala Val Asn Gly Thr Arg Asn Val Leu Leu Ser Ile 100 105 110
- Lys Asp Asn Leu Pro Asn Val Lys Arg Phe Val Tyr Thr Ser Ser Leu
- Ala Ala Val Arg Thr Glu Gly Ala Gly Tyr Ser Ala Asp Glu Val Val
- Thr Glu Asp Ser Trp Asn Asn Ile Ala Leu Lys Asp Ala Thr Lys Asp 145 150 155 160
- Glu Gly Thr Ala Tyr Glu Ala Ser Lys Thr Tyr Gly Glu Lys Glu Val 165 170 175
- Trp Asn Phe Phe Glu Lys Thr Lys Asn Val Asn Phe Asp Phe Ala Ile 180 185 190
- Ile Asn Pro Val Tyr Val Phe Gly Pro Gln Leu Phe Glu Glu Tyr Val 195 200 205
- Thr Asp Lys Leu Asn Phe Ser Ser Glu Ile Ile Asn Ser Ile Ile Lys 210 215 220
- Gly Glu Lys Lys Glu Ile Glu Gly Tyr Glu Ile Asp Val Arg Asp Ile 225 230 235 240
- Ala Arg Ala His Ile Ser Ala Val Glu Asn Pro Ala Thr Thr Arg Gln 245 250 255

- 5 -

Arg Leu Ile Pro Ala Val Ala Pro Tyr Asn Gln Gln Thr Ile Leu Asp 260 265 270

Val Leu Asn Glu Asn Phe Pro Glu Leu Lys Gly Lys Ile Asp Val Gly
275 280 285

Lys Pro Gly Ser Gln Asn Glu Phe Ile Lys Lys Tyr Tyr Lys Leu Asp 290 295 300

Asn Ser Lys Thr Lys Lys Val Leu Gly Phe Glu Phe Ile Ser Gln Glu 305 310 315 320

Gln Thr Ile Lys Asp Ala Ala Gln Ile Leu Ser Val Lys Asn Gly
325 330 335

Lys Lys

(2) INFORMATION FOR SEQ ID NO:3:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1271 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: mRNA

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

UGAAUGGUUA UUUUAGCAAU UGCUGUGUGA GGCACUGACC UAAAGAUGUG UAUAAAUAGU 60 GGGACUGUGU ACUCAUGAGG AUCAAUACAU GUAUAAACUU ACCAUACUUU CACACAAGUC 120 AACUUAGAAU CAAUCAAUCA AUCAAUUAAU CAAGCUAUAC AAUAUGACAA AAGUCUUCGU 180 AACAGGUGCC AACGGAUUCG UUGCUCAACA CGUCGUUCAU CAACUAUUAG AAAAGAACUA 240 UACAGUGGUU GGAUCUGUCC GUUCAACUGA GAAAGGUGAU AAAUUAGCUA AAUUGCUAAA 300 CAAUCCAAAA UUUUCAUAUG AGAUUAUUAA AGAUAUGGUC AAUUCGAGAG AUGAAUUCGA 360 UAAGGCUUUA CAAAAACAUU CAGAUGUUGA AAUUGUCUUA CAUACUGCUU CACCAGUCUU 420 CCCAGGUGGU AUUAAAGAUG UUGAAAAAGA AAUGAUCCAA CCAGCUGUUA AUGGUACUAG 480 AAAUGUCUUG UUAUCAAUCA AGGAUAACUU ACCAAAUGUC AAGAGAUUUG UUUACACUUC 540

UUCAUUAGCU	GCUGUCCGUA	CUGAAGGUGC	UGGUUAUAGU	GCAGACGAAG	UUGUCACCGA	600
AGAUUCUUGG	AACAAUAUUG	CAUUGAAAGA	UGCCACCAAG	GAUGAAGGUA	CAGCUUAUGA	660
GGCUUCCAAG	ACAUAUGGUG	AAAAAGAAGU	UUGGAAUUUC	UUCGAAAAA	CUAAAAAUGU	720
UAAUUUCGAU	UUUGCCAUCA	UCAACCCAGU	UUAUGUCUUU	GGUCCUCAAU	UAUUUGAAGA	780
AUACGUUACU	GAUAAAUUGA	ACUUUUCCAG	UGAAAUCAUU	AAUAGUAUAA	UAAAAGGUGA	840
AAAGAAGGAA	AUUGAAGGUU	AUGAAAUUGA	UGUUAGAGAU	AUUGCAAGAG	CUCAUAUCUC	900
UGCUGUUGAA	AAUCCAGCAA	CUACACGUCA	AAGAUUAAUU	CCAGCAGUUG	CACCAUACAA	960
UCAACAAACU	AUCUUGGAUG	UUUUGAAUGA	AAACUUCCCA	GAAUUGAAAG	GUAAAAUCGA	1020
UGUUGGGAAA	CCAGGUUCUC	AAAAUGAAUU	AAAAAUUAU	UAAAUAUUAU	UAGAUAACUC	1080
AAAGACCAAA	AAAGUUUUAG	GUUUUGAAUU	CAUUUCCCAA	GAGCAAACAA	UCAAAGAUGC	1140
UGCUGCUCAA	AUCUUGUCCG	UUAAAAAUGG	AAUAAAAAA	GUGAACUAGA	CCUGUCACUA	1200
UCAGAUUAUU	AGAGUUCUGU	AUAGAUUAAA	GUGUGAAAAU	GUAUUAGAAU	CAUAAUUUUA	1260
UAAUUAUGCC	U					1271

(2) INFORMATION FOR SEQ ID NO:4:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1032 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (iii) HYPOTHETICAL: NO
 - (iv) ANTI-SENSE: NO
 - (ix) FEATURE:
 - (A) NAME/KEY: CDS
 - (B) LOCATION: 1..1032
 - (D) OTHER INFORMATION: S.cerevisiae YDR541c
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

ATG Met 1	TCT Ser	Asn	Thr	Val	Leu	Val	Ser	Gly	Ala	Ser	GGT Gly	Phe	Ile	Ala	TTG Leu	48
CAT	ATC	CTG	TCA	CAA	TTG	TTA	AAA	CAA	GAT	TAT	AAG	GTT	ATT	GGA	ACT	96

His Ile Leu Ser Gln Leu Leu Lys Gln Asp Tyr Lys Val Ile Gly Thr
20 25 30

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GTG Val	AGA Arg	TCC Ser 35	CAT His	GAA Glu	AAA Lys	GAA Glu	GCA Ala 40	AAA Lys	TTG Leu	CTA Leu	AGA Arg	CAA Gln 45	TTT Phe	CAA Gln	CAT His	144
AAC Asn	CCT Pro 50	AAT Asn	TTA Leu	ACT Thr	TTA Leu	GAA Glu 55	ATT Ile	GTT Val	CCG Pro	GAC Asp	ATT Ile 60	TCT Ser	CAT His	CCA Pro	AAT Asn	192
GCT Ala 65	TTC Phe	GAT Asp	AAG Lys	GTT Val	CTG Leu 70	CAG Gln	AAA Lys	CGT Arg	GGA Gly	CGT Arg 75	GAG Glu	ATT Ile	AGG Arg	TAT Tyr	GTT Val 80	240
CTA Leu	CAC His	ACG Thr	GCC Ala	TCT Ser 85	CCT Pro	TTT Phe	CAT His	TAT Tyr	GAT Asp 90	ACT Thr	ACC Thr	GAA Glu	TAT Tyr	GAA Glu 95	AAA Lys	288
GAC Asp	TTA Leu	TTG Leu	ATT Ile 100	CCC Pro	GCG Ala	TTA Leu	GAA Glu	GGT Gly 105	ACA Thr	AAA Lys	AAC Asn	ATC Ile	CTA Leu 110	TAA Asn	TCT Ser	336
Ile	AAG Lys	Lys 115	Tyr	Ala	Ala	Asp	Thr 120	Val	Glu	Arg	Val	Val 125	Val	Thr	Ser	384
Ser	TGT Cys 130	Thr	Ala	Ile	Ile	Thr 135	Leu	Ala	Lys	Met	Asp 140	Asp	Pro	Ser	Val	432
Val 145	TTT	Thr	Glu	Glu	Ser 150	Trp	Asn	Glu	Ala	Thr 155	Trp	Glu	Ser	Суз	Gln 160	480
Ile	GAT Asp	Gly	Ile	Asn 165	Ala	Tyr	Phe	Ala	Ser 170	Lys	Lys	Phe	Ala	Glu 175	Lys	528
Ala	GCC Ala	Trp	Glu 180	Phe	Thr	Lys	Glu	Asn 185	Glu	Asp	His	Ile	Lys 190	Phe	Lys	576
Leu	ACA Thr	Thr 195	Val	Asn	Pro	Ser	Leu 200	Leu	Phe	Gly	Pro	Gln 205	Leu	Phe	qaA	624
Glu	GAT Asp 210	Val	His	Gly	His	Leu 215	Asn	Thr	Ser	Cys	Glu 220	Met	Ile	Asn	Gly	672
Leu 225		His	Thr	Pro	Val 230	Asn	Ala	Ser	Val	Pro 235	Asp	Pire	His	Ser	1le 240	720
TTT Phe	ATT	GAT Asp	GTA Val	AGG	GAT Asp	GTG Val	GCC	CTA Leu	GCT Ala	CAT His	CTG Leu	TAT Tyr	GCT Ala	TTC Phe	CAG Gln	768

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							_							
		245					250					255		
AAG GAA A Lys Glu A	AAT ACC Asn Thr 260	GCG Ala	GGT Gly	AAA Lys	AGA Arg	TTA Leu 265	GTG Val	GTA Val	ACT Thr	AAC Asn	GGT Gly 270	AAA Lys	TTT Phe	816
GGA AAC G	CAA GAT Gln Asp 275	ATC Ile	CTG Leu	GAT Asp	ATT Ile 280	TTG Leu	AAC Asn	GAA Glu	GAT Asp	TTT Phe 285	CCA Pro	CAA Gln	TTA Leu	864
AGA GGT (Arg Gly I 290	CTC ATT Leu Ile	CCT Pro	TTG Leu	GGT Gly 295	AAG Lys	CCT Pro	GGC Gly	ACA Thr	GGT Gly 300	GAT Asp	CAA Gln	GTC Val	ATT Ile	912
GAC CGC (Asp Arg (GGT TCA Gly Ser	ACT Thr	ACA Thr 310	GAT Asp	AAT Asn	AGT Ser	GCA Ala	ACG Thr 315	AGG Arg	AAA Lys	ATA Ile	CTT Leu	GGC Gly 320	960
TTT GAG	TTC AGA Phe Arg	AGT Ser 325	TTA Leu	CAC His	GAA Glu	AGT Ser	GTC Val 330	CAT His	GAT Asp	ACT Thr	GCT Ala	GCC Ala 335	CAA Gln	1008
ATT TTG A														1032
(2) INFO	RMATION	FOR	SEQ	ID N	iO: 5:	:								
(:	(B	ENCE) LEN) TYI) TOI	NGTH: PE: a	344 min	am:	ino a id		3						•
(i	i) MOLE	CULE	TYPE	E: p	rote:	in								
(x	i) SEQU	ENCE	DESC	RIP	rion	: SE(O ID	NO:	5:					
Met Ser	Asn Thr	Val 5	Leu	Val	Ser	Gly	Ala 10	Ser	Gly	Phe	Ile	Ala 15	Leu	
His Ile	Leu Ser 20	Gln	Leu	Leu	Lys	Gln 25	Asp	Tyr	Lys	Val	Ile 30	Gly	Thr	
Val Arg	Ser His 35	Glu	Lys	Glu	Ala 40	Lys	Leu	Leu	Arg	Gln 45	Phe	Gln	His	
Asn Pro 50	Asn Leu	Thr	Leu	Glu 55	Ile	Val	Pro	Asp	Ile 60	Ser	His	Pro	Asn	
Ala Phe 65	Asp Lys	Val	Leu 70	Gln	Lys	Arg	Gly	Arg 75	Glu	Ile	Arg	Tyr	Val 80	
Leu His														

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Asp	Leu	Leu	Ile 100	Pro	Ala	Leu	Glu	Gly 105	Thr	Lys	Asn	Ile	Leu 110	Asn	Ser
Ile	Lys	Lys 115	Tyr	Ala	Ala	Asp	Thr 120	Val	Glu	Arg	Val	Val 125	Val	Thr	Ser
Ser	Сув 130	Thr	Ala	Ile	Ile	Thr 135	Leu	Ala	Lys	Met	Asp 140	Asp	Pro	Ser	Val
Val 145	Phe	Thr	Glu	Glu	Ser 150	Trp	Asn	Glu	Ala	Thr 155	Trp	Glu	Ser	Cys	Gln 160
Ile	Asp	Gly	Ile	Asn 165	Ala	Tyr	Phe	Ala	Ser 170	Lys	Lys	Phe	Ala	Glu 175	Lys
Ala	Ala	Trp	Glu 180	Phe	Thr	Lys	Glu	Asn 185	Glu	Asp	His	Ile	Lys 190	Phe	Lys
Leu	Thr	Thr 195	Val	Asn	Pro	Ser	Leu 200	Leu	Phe	Gly	Pro	Gln 205	Leu	Phe	Asp
Glu	Asp 210	Val	His	Gly	His	Leu 215	Asn	Thr	Ser	Cys	Glu 220	Met	Ile	naA	Gly
Leu 225	Ile	His	Thr	Pro	Val 230	Asn	Ala	Ser	Val	Pro 235	Asp	Phe	His	Ser	Ile 240
Phe	Ile	Asp	Val	Arg 245	Asp	Val	Ala	Leu	Ala 250	His	Leu	Tyr	Ala	Phe 255	Gln
Lys	Glu	Asn	Thr 260	Ala	Gly	Lys	Arg	Leu 265	Val	Val	Thr	Asn	Gly 270	Lys	Phe
Gly	Asn	Gln 275	Asp	Ile	Leu	Asp	Ile 280	Leu	Asn	Glu	Asp	Phe 285	Pro	Gln	Leu
Arg	Gly 290	Leu	Ile	Pro	Leu	Gly 295	Lys	Pro	Gly	Thr	Gly 300	Asp	Gln	Val	Ile
Asp 305	_	Gly	Ser	Thr	Thr 310	Asp	Asn	Ser	Ala	Thr 315	Arg	Lys	Ile	Leu	Gly 320
Phe	Glu	Phe	Arg	Ser 325	Leu	His	Glu	Ser	Val 330	His	Asp	Thr	Ala	Ala 335	Gln

(2) INFORMATION FOR SEQ ID NO:6:

Ile Leu Lys Lys Glu Asn Arg Leu 340

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1032 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: mRNA

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

AUGUCUAAUA CAGUUCUAGU UUCUGGCGCU UCAGGUUUUA UUGCCUUGCA UAUCCUGUCA	60
CAAUUGUUAA AACAAGAUUA UAAGGUUAUU GGAACUGUGA GAUCCCAUGA AAAAGAAGCA	120
AAAUUGCUAA GACAAUUUCA ACAUAACCCU AAUUUAACUU UAGAAAUUGU UCCGGACAUU	180
UCUCAUCCAA AUGCUUUCGA UAAGGUUCUG CAGAAACGUG GACGUGAGAU UAGGUAUGUU	240
CUACACACGG CCUCUCCUUU UCAUUAUGAU ACUACCGAAU AUGAAAAAGA CUUAUUGAUU	300
CCCGCGUUAG AAGGUACAAA AAACAUCCUA AAUUCUAUCA AGAAAUAUGC AGCAGACACU	360
GUAGAGCGUG UUGUUGUGAC UUCUUCUUGU ACUGCUAUUA UAACCCUUGC AAAGAUGGAC	420
GAUCCCAGUG UGGUUUUUAC AGAAGAGAGU UGGAACGAAG CAACCUGGGA AAGCUGUCAA	480
AUUGAUGGGA UAAAUGCUUA CUUUGCAUCC AAGAAGUUUG CUGAAAAGGC UGCCUGGGAG	540
UUCACAAAAG AGAAUGAAGA UCACAUCAAA UUCAAACUAA CAACAGUCAA CCCUUCUCUU	600
CUUUUUGGUC CUCAACUUUU CGAUGAAGAU GUGCAUGGCC AUUUGAAUAC UUCUUGCGAA	660
AUGAUCAAUG GCCUAAUUCA UACCCCAGUA AAUGCCAGUG UUCCUGAUUU UCAUUCCAUU	720
UUUAUUGAUG UAAGGGAUGU GGCCCUAGCU CAUCUGUAUG CUUUCCAGAA GGAAAAUACC	780
GCGGGUAAAA GAUUAGUGGU AACUAACGGU AAAUUUGGAA ACCAAGAUAU CCUGGAUAUU	840
UUGAACGAAG AUUUUCCACA AUUAAGAGGU CUCAUUCCUU UGGGUAAGCC UGGCACAGGU	900
GAUCAAGUCA UUGACCGCGG UUCAACUACA GAUAAUAGUG CAACGAGGAA AAUACUUGGC	960
UUUGAGUUCA GAAGUUUACA CGAAAGUGUC CAUGAUACUG CUGCCCAAAU UUUGAAGAAG	1020
GAGAACAGAU UA	1032

(2) INFORMATION FOR SEQ ID NO:7:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1029 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single

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		(E) TO	POLC	GY:	line	ear									
	(ii)	MOL	ECUL	E TY	PE:	DNA	(gen	omic	:)		•					
- (iii)	нуг	OTHE	TICA	L: N	Ю										
	/21	74 8 17 17	'I-SE	NEF.	NΩ											
	(1V)	ANI	1-5E	.acn.	NO											
	(ix)	(<i>p</i>	ATURE A) NA B) LC D) OI	ME/K	ON:	11	.026 PION:	s.c	erev	risiā	ne YC)L151	.w			
	(xi)	SEC	UENC	E DE	SCRI	PTIC	N: S	EQ 1	D NC):7:						
ATG	TCA	GTT	TTC	GTT	TCA	GGT	GCT	AAC	GGG	TTC	ATT	GCC	CAA	CAC	ATT	48
Met	Ser	Val	Phe		Ser	Gly	Ala	Asn		Phe	Ile	Ala	Gln	His 15	Ile	
1				5					10							
GTC	GAT	CTC	CTG	TTG	AAG	GAA	GAC	TAT	AAG	GTC	ATC	GGT	TCT	GCC	AGA	96
Val	Asp	Leu.	Leu 20	Leu	Lys	Glu	Ąsp	Tyr 25	Lys	Val	Пе	GIY	ser 30	Ala	Arg	
AGT	CAA	GAA	AAG	GCC	GAG	AAT	TTA	ACG	GAG	GCC	TTT	GGT	AAC	AAC	CCA	144
Ser	Gln	G1u 35	Lys	Ala	GIU	ASII	Leu 40	THE	GIU	мта	FHE	45	LOII	ABII	110	
												~~~	~~~		menon.	100
AAA	TTC	TCC	ATG	GAA	GTT	GTC Val	CCA Pro	GAC	ATA Ile	Ser	LVS	Leu	Asp	Ala	Phe	192
гÀг	50	Ser	Mec	Giu	Val	55	110	p			60		•			
		·				a. a	GGC	n n C	CATT	እጥሮ	AAG	አጥል	بلبيلت	CTD	_ር ልሞ	240
GAC	CAT	GTT Val	Phe	Gln	LVS	His	Gly	Lys	Asp	Ile	Lys	Ile	Val	Leu	His	210
65					70		-	-	_	75					80	
3.00	000	ui Cati	CCA	יייייי	TGC	Jerter	GAT	АТС	ACT	GAC	AGT	GAA	CGC	GAT	TTA	288
Thr	Ala	Ser	Pro	Phe	Cys	Phe	Asp	Ile	Thr	Asp	Ser	Glu	Arg	Asp	Leu	
				85					90					95		•
ጥጥል	יויינע	רירייוי	GCT	GTG	AAC	GGT	GTT	AAG	GGA	ATT	CTC	CAC	TCA	ATT	AAA	336
Leu	Ile	Pro	Ala	Val	Asn	Gly	Val	Lys	Gly	Ile	Leu	His	Ser	Ile	Lys	
			100					105					110			
AAA	TAC	GCC	GCT	GAT	TCT	GTA	GAA	CGT	GTA	GTT	CTC	ACC	TCT	TCT	TAT	384
Lys	Tyr	Ala	Ala	Asp	Ser	Val	Glu	Arg	Val	Val	Leu	Thr	Ser	Ser	Tyr	
,		115					120					125				
GCA	GCT	GTG	TTC	GAT	ATG	GCA	AAA	GAA	AAC	GAT	AAG	TCT	TTA	ACA	TTT	432
Ala	Ala	Val	Phe	Asp	Met	Ala	Lys	Glu	Asn	Asp	Lys	Ser	Leu	Thr	Phe	
	130					135					140					
AAC	GAA	GAA	TCC	TGG	AAC	CCA	GCT	ACC	TGG	GAG	AGT	TGC	CAA	AGT	GAC	480
Acn	Glu	Glu	Ser	Trans	Asn	Pro	Ala	Thr	Trp	Glu	Ser	Cys	Gln	Ser	Asp	-

Asn Glu Glu Ser Trp Asn Pro Ala Thr Trp Glu Ser Cys Gln Ser Asp

155

150

145

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CCA Pro	GTT Val	AAC Asn	GCC Ala	TAC Tyr 165	TGT Cys	GGT Gly	TCT Ser	AAG Lys	AAG Lys 170	TTT Phe	GCT Ala	GAA Glu	AAA Lys	GCA Ala 175	GCT Ala	528
TGG Trp	GAA Glu	TTT Phe	CTA Leu 180	GAG Glu	GAG Glu	AAT Asn	AGA Arg	GAC Asp 185	TCT Ser	GTA Val	AAA Lys	TTC Phe	GAA Glu 190	TTA Leu	ACT Thr	576
GCC Ala	GTT Val	AAC Asn 195	CCA Pro	GTT Val	TAC Tyr	GTT Val	TTT Phe 200	GGT Gly	CCG Pro	CAA Gln	ATG Met	TTT Phe 205	GAC Asp	AAA Lys	GAT Asp	624
GTG Val	AAA Lys 210	AAA Lys	CAC His	TTG Leu	AAC Asn	ACA Thr 215	TCT Ser	TGC Cys	GAA Glu	CTC Leu	GTC Val 220	AAC Asn	AGC Ser	TTG Leu	ATG Met	672
CAT His 225	TTA Leu	TCA Ser	CCA Pro	GAG Glu	GAC Asp 230	AAG Lys	ATA Ile	CCG Pro	GAA Glu	CTA Leu 235	TTT Phe	GGT Gly	GGA Gly	TAC Tyr	ATT Ile 240	720
GAT Asp	GTT Val	CGT Arg	GAT Asp	GTT Val 245	GCA Ala	AAG Lys	GCT Ala	CAT His	TTA Leu 250	GTT Val	GCC Ala	TTC Phe	CAA Gln	AAG Lys 255	AGG Arg	768
GAA Glu	ACA Thr	ATT Ile	GGT Gly 260	CAA Gln	AGA Arg	CTA Leu	ATC Ile	GTA Val 265	TCG Ser	GAG Glu	GCC Ala	AGA Arg	TTT Phe 270	ACT Thr	ATG Met	816
CAG Gln	GAT Asp	GTT Val 275	CTC Leu	GAT Asp	ATC Ile	CIT Leu	AAC Asn 280	GAA Glu	GAC Asp	TTC Phe	CCT Pro	GTT Val 285	CTA Leu	AAA Lys	GGC	864
AAT Asn	ATT Ile 290	CCA Pro	GTG Val	GGG Gly	AAA Lys	CCA Pro 295	GGT Gly	TCT Ser	GGT Gly	GCT Ala	ACC Thr 300	CAT His	AAC Asn	ACC Thr	CTT Leu	912
GGT Gly 305	GCT Ala	ACT Thr	CTT Leu	GAT Asp	AAT Asn 310	AAA Lys	AAG Lys	AGT Ser	AAG Lys	AAA Lys 315	TTG Leu	TTA Leu	GGT Gly	TTC Phe	AAG Lys 320	960
TTC Phe	AGG Arg	AAC Asn	TTG Leu	AAA Lys 325	Glu	ACC Thr	ATT	GAC Asp	GAC Asp 330	ACT Thr	GCC Ala	TCC Ser	CAA Gln	ATT Ile 335	TTA Leu	1008
				AGA Arg	_	TAA										1029

#### (2) INFORMATION FOR SEQ ID NO:8:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 342 amino acids
  - (B) TYPE: amino acid

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- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:
- Met Ser Val Phe Val Ser Gly Ala Asn Gly Phe Ile Ala Gln His Ile
- Val Asp Leu Leu Leu Lys Glu Asp Tyr Lys Val Ile Gly Ser Ala Arg 20 25 30
- Ser Gln Glu Lys Ala Glu Asn Leu Thr Glu Ala Phe Gly Asn Asn Pro
- Lys Phe Ser Met Glu Val Val Pro Asp Ile Ser Lys Leu Asp Ala Phe
  50 55 60
- Asp His Val Phe Gln Lys His Gly Lys Asp Ile Lys Ile Val Leu His 65 70 75 80
- Thr Ala Ser Pro Phe Cys Phe Asp Ile Thr Asp Ser Glu Arg Asp Leu 85 90 95
- Leu Ile Pro Ala Val Asn Gly Val Lys Gly Ile Leu His Ser Ile Lys
  100 105 110
- Lys Tyr Ala Ala Asp Ser Val Glu Arg Val Val Leu Thr Ser Ser Tyr 115 120 125
- Ala Ala Val Phe Asp Met Ala Lys Glu Asn Asp Lys Ser Leu Thr Phe 130 135 140
- Asn Glu Glu Ser Trp Asn Pro Ala Thr Trp Glu Ser Cys Gln Ser Asp 145 150 155 160
- Pro Val Asn Ala Tyr Cys Gly Ser Lys Lys Phe Ala Glu Lys Ala Ala 165 170 175
- Trp Glu Phe Leu Glu Glu Asn Arg Asp Ser Val Lys Phe Glu Leu Thr 180 185 190
- Ala Val Asn Pro Val Tyr Val Phe Gly Pro Gln Met Phe Asp Lys Asp 195 200 205
- Val Lys Lys His Leu Asn Thr Ser Cys Glu Leu Val Asn Ser Leu Met 210 215 220
- His Leu Ser Pro Glu Asp Lys Ile Pro Glu Leu Phe Gly Gly Tyr Ile 225 230 235 240
- Asp Val Arg Asp Val Ala Lys Ala His Leu Val Ala Phe Gln Lys Arg 245 250 255
- Glu Thr Ile Gly Gln Arg Leu Ile Val Ser Glu Ala Arg Phe Thr Met

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 Gln Asp Val 275
 Leu Asp 275
 Ile Leu Asp 280
 Glu Asp Glu Asp 280
 Phe Pro Val 285
 Leu Lys Gly 285

 Asn Ile 290
 Pro Val Gly Lys Pro 295
 Ser Gly Ala Thr His Asn Thr Leu 300
 Asn Lys Lys Ser Lys Lys Lys Leu Leu Gly Phe Lys 305

Phe Arg Asn Leu Lys Glu Thr Ile Asp Asp Thr Ala Ser Gln Ile Leu 325 330 335

Lys Phe Glu Gly Arg Ile 340

#### (2) INFORMATION FOR SEQ ID NO:9:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 1026 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: mRNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO

#### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

AUGUCAGUUU	UCGUUUCAGG	UGCUAACGGG	UUCAUUGCCC	AACACAUUGU	CGAUCUCCUG	60
UUGAAGGAAG	ACUAUAAGGU	CAUCGGUUCU	GCCAGAAGUC	AAGAAAAGGC	CGAGAAUUUA	120-
ACGGAGGCCU	UUGGUAACAA	CCCAAAAUUC	UCCAUGGAAG	UUGUCCCAGA	CAUAUCUAAG	180
CUGGACGCAU	UUGACCAUGU	UUUCCAAAAG	CACGGCAAGG	AUAUCAAGAU	AGUUCUACAU	240
ACGGCCUCUC	CAUUCUGCUU	UGAUAUCACU	GACAGUGAAC	GCGAUUUAUU	AAUUCCUGCU	300
GUGAACGGUG	UUAAGGGAAU	UCUCCACUCA	UAAAAAUUA	ACGCCGCUGA	UUCUGUAGAA	360
CGUGUAGUUC	UCACCUCUUC	UUAUGCAGCU	GUGUUCGAUA	UGGCAAAAGA	AAACGAUAAG	420
UCUUUAACAU	UUAACGAAGA	AUCCUGGAAC	CCAGCUACCU	GGGAGAGUUG	CCAAAGUGAC	480
CCAGUUAACG	CCUACUGUGG	UUCUAAGAAG	UUUGCUGAAA	AAGCAGCUUG	GGAAUUUCUA	540
GAGGAGAAUA	GAGACUCUGU	AAAAUUCGAA	UUAACUGCCG	UUAACCCAGU	UUACGUUUUU	600

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GGUCCGCAAA UGUUUGACAA AGAUGUGAAA AAACACUUGA ACACAUCUUG CGAACUCGUC	660
AACAGCUUGA UGCAUUUAUC ACCAGAGGAC AAGAUACCGG AACUAUUUGG UGGAUACAUU	720
GAUGUUCGUG AUGUUGCAAA GGCUCAUUUA GUUGCCUUCC AAAAGAGGGGA AACAAUUGGU	780
CAAAGACUAA UCGUAUCGGA GGCCAGAUUU ACUAUGCAGG AUGUUCUCGA UAUCCUUAAC	840
GAAGACUUCC CUGUUCUAAA AGGCAAUAUU CCAGUGGGGA AACCAGGUUC UGGUGCUACC	900
CAUAACACCC UUGGUGCUAC UCUUGAUAAU AAAAAGAGUA AGAAAUUGUU AGGUUUCAAG	960
UUCAGGAACU UGAAAGAGAC CAUUGACGAC ACUGCCUCCC AAAUUUUAAA AUUUGAGGGC	1020
AGAAUA	1026
(2) INFORMATION FOR SEQ ID NO:10:	
(i) SEQUENCE CHARACTERISTICS:	
(a) renown. 1041 bace pairs	

- (A) LENGTH: 1041 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (ix) FEATURE:
  - (A) NAME/KEY: CDS
  - (B) LOCATION: 1..1041
  - (D) OTHER INFORMATION: S. cerevisiae YGL157W
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

ATG Met 1	ACT Thr	ACT Thr	GAT Asp	ACC Thr 5	ACT Thr	GTT Val	TTC Phe	GTT Val	TCT Ser 10	GGC Gly	GCA Ala	ACC Thr	GGT Gly	TTC Phe 15	ATT Ile	48
										GCT Ala						96
										GGC Gly						144
										GTG Val						192

55 60

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													GAA Glu		AAG Lys 80	240
													ACC Thr			288
													TCT Ser 110			336
													GTT Val			384
													AAC Asn			432
Asp 145	Leu	Val	Ile	Thr	Glu 150	Glu	Ser	Trp	Asn	Lys 155	qaA	Thr	TGG Trp	Asp	Ser 160	480
Cys	Gln	Ala	Asn	Ala 165	Val	Ala	Ala	Tyr	Cys 170	Gly	Ser	Lys	AAG Lys	Phe 175	Ala	528
													AGT Ser 190			576
	_		180													
Phe	Thr	Leu 195	TCC Ser	Thr	Ile	Asn	Pro 200	Gly	Phe	Val	Phe	Gly 205	CCT Pro	Gln	Met	624
Phe TTT Phe	Thr GCA Ala 210	Leu 195 GAT Asp	TCC Ser TCG Ser	Thr CTA Leu	Ile AAA Lys	CAT His 215	Pro 200 GGC Gly	Gly ATA Ile	Phe AAT Asn	Val ACC Thr	TCC Ser 220	Gly 205 TCA Ser	Pro GGG Gly	Gln ATC Ile	Met GTA Val	624 672
TTT Phe TCT Ser 225	GCA Ala 210 GAG Glu	Leu 195 GAT Asp TTA Leu	TCC Ser TCG Ser ATT Ile	Thr CTA Leu CAT His	AAA Lys TCC Ser 230	CAT His 215 AAG Lys	Pro 200 GGC Gly GTA Val	Gly ATA Ile GGT Gly	AAT Asn GGA Gly	ACC Thr GAA Glu 235	TCC Ser 220 TTT Phe	Gly 205 TCA Ser TAT Tyr	GGG Gly AAT Asn	Gln ATC Ile TAC Tyr	Met GTA Val TGT Cys 240	
TTT Phe TCT Ser 225 GGC Gly	GCA Ala 210 GAG Glu CCA Pro	Leu 195 GAT Asp TTA Leu TTT Phe	TCC Ser TCG Ser ATT Ile	CTA Leu CAT His GAC Asp 245	AAA Lys TCC Ser 230 GTG Val	CAT His 215 AAG Lys CGT Arg	Pro 200 GGC Gly GTA Val GAC Asp	ATA Ile GGT Gly GTT Val	AAT Asn GGA Gly TCT Ser 250	ACC Thr GAA Glu 235 AAA Lys	TCC Ser 220 TTT Phe GCC Ala	Gly 205 TCA Ser TAT Tyr CAC His	GGG Gly AAT Asn CTA Leu	Gln ATC Ile TAC Tyr GTT Val 255	Met GTA Val TGT Cys 240 GCA Ala	672
TTT Phe TCT Ser 225 GGC Gly ATT Ile	GCA Ala 210 GAG Glu CCA Pro	Leu 195 GAT Asp TTA Leu TTT Phe	TCC Ser TCG Ser ATT Ile ATT Ile	CTA Leu CAT His GAC Asp 245 GAA Glu	AAA Lys TCC Ser 230 GTG Val	CAT His 215 AAG Lys CGT Arg	Pro 200 GGC Gly GTA Val GAC Asp	Gly ATA Ile GGT Gly GTT Val CAA Gln 265	AAT Asn GGA Gly TCT Ser 250 AGA Arg	ACC Thr GAA Glu 235 AAA Lys	TCC Ser 220 TTT Phe GCC Ala GTA Val	Gly 205 TCA Ser TAT Tyr CAC His	GGG Gly AAT Asn	Gln ATC Ile TAC Tyr GTT Val 255 GAA Glu	Met GTA Val TGT Cys 240 GCA Ala GGT Gly	672 720

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			GGC Gly													912
			AAA Lys													960
			TTC Phe													1008
			ATG Met 340													1041
(2)	INFO	ORMA'	rion	FOR	SEQ	ID 1	NO:11	L:								
		(i) \$	(B)	LEN TYP	NGTH:		am:	ino a	: acids	5						
	(:	ii) 1	MOLE													
	(2	ki) 8	SEQUI	BNCE	DESC	CRIP	rion:	: SE(	Q ID	NO:1	L <b>1</b> :					
Met 1			SEQUI Asp									Thr	Gly	Phe 15	Ile	
1	Thr	Thr		Thr 5	Thr	Val	Phe	Val	Ser 10	Gly	Ala			15		
1 Ala	Thr	Thr	Asp	Thr 5 Met	Thr Asn	Val Asp	Phe Leu	Val Leu 25	Ser 10 Lys	Gly Ala	Ala Gly	Tyr	Thr 30	15 Val	Ile	
1 Ala Gly	Thr Leu Ser	Thr His Gly 35	Asp Ile 20	Thr 5 Met Ser	Thr Asn Gln	Val Asp Glu	Phe Leu Lys 40	Val Leu 25 Asn	Ser 10 Lys Asp	Gly Ala Gly	Ala Gly Leu	Tyr Leu 45	Thr 30 Lys	15 Val Lys	Ile Phe	
1 Ala Gly Asn	Thr Leu Ser Asn 50	Thr His Gly 35 Asn	Asp Ile 20 Arg	Thr 5 Met Ser	Thr Asn Gln Leu	Val Asp Glu Ser 55	Phe Leu Lys 40 Met	Val Leu 25 Asn Glu	Ser 10 Lys Asp	Gly Ala Gly Val	Ala Gly Leu Glu 60	Tyr Leu 45 Asp	Thr 30 Lys Ile	15 Val Lys Ala	Ile Phe Ala	
Ala Gly Asn Pro 65	Thr Leu Ser Asn 50 Asn	Thr His Gly 35 Asn	Asp Ile 20 Arg	Thr 5 Met Ser Lys	Thr Asn Gln Leu Glu 70	Val Asp Glu Ser 55	Phe Leu Lys 40 Met	Val Leu 25 Asn Glu Lys	Ser 10 Lys Asp Ile	Gly Ala Gly Val His 75	Ala Gly Leu Glu 60 Gly	Tyr Leu 45 Asp	Thr 30 Lys Ile Glu	15 Val Lys Ala Ile	Ile Phe Ala Lys 80	
1 Ala Gly Asn Pro 65	Thr Leu Ser Asn 50 Asn Val	Thr His Gly 35 Asn Ala	Ile 20 Arg Pro	Thr 5 Met Ser Lys Asp Thr 85	Thr Asn Gln Leu Glu 70 Ala	Val Asp Glu Ser 55 Val	Phe Leu Lys 40 Met Phe	Leu 25 Asn Glu Lys	Ser 10 Lys Asp Ile Lys His 90	Gly Ala Gly Val His 75	Ala Gly Leu Glu 60 Gly Glu	Tyr Leu 45 Asp Lys	Thr 30 Lys Ile Glu	Val Lys Ala Ile Asn 95	Ile Phe Ala Lys 80 Phe	

Thr Ser Ser Thr Ala Ala Leu Val Thr Pro Thr Asp Met Asn Lys Gly

140

135

130

Asp 145	Leu	Val	Ile	Thr	Glu 150	Glu	Ser	Trp	Asn	Lys 155	Asp	Thr	Trp	Asp	Ser 160
Сув	Gln	Ala	Asn	Ala 165	Val	Ala	Ala	Tyr	Cys 170	Gly	Ser	Lys	Lys	Phe 175	Ala
Glu	Lys	Thr	Ala 180	Trp	Glu	Phe	Leu	Lys 185	Glu	Asn	Lys	Ser	Ser 190	Val	ГÀЗ
Phe	Thr	Leu 195	Ser	Thr	Ile	Asn	Pro 200	Gly	Phe	Val	Phe	Gly 205	Pro	Gln	Met
Phe	Ala 210	Asp	Ser	Leu	Lys	His 215	Gly	Ile	Asn	Thr	Ser 220	Ser	Gly	Ile	Val
Ser 225	Glu	Leu	Ile	His	Ser 230	Lys	Val	Gly	Gly	Glu 235	Phe	Tyr	Asn	Tyr	Cys 240
Gly	Pro	Phe	Ile	Asp 245	Val	Arg	Asp	Val	Ser 250	Lys	Ala	His	Leu	<b>Val</b> 255	Ala
Ile	Glu	Lys	Pro 260	Glu	Cys	Thr	Gly	Gln 265	Arg	Leu	Val	Leu	Ser 270	Glu	Gly
Leu	Phe	Cys 275	Сув	Gln	Glu	Ile	Val 280	Asp	Ile	Leu	Asn	Glu 285	Glu	Phe	Pro
Gln	Leu 290	Lys	Gly	Lys	Ile	Ala 295	Thr	Gly	Glu	Pro	Ala 300	Thr	Gly	Pro	Ser
Phe 305		Glu	Lys	Asn	Ser 310	Cys	Lys	Phe	Asp	Asn 315	Ser	Lys	Thr	Lys	Lys 320
Leu	Leu	Gly	Phe	Gln	Phe	Tyr	Asn	Leu	Lys	Asp	Cys	Ile	Val	Asp	Thr

345

(2) INFORMATION FOR SEQ ID NO:12:

340

325

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 1041 base pairs
  - (B) TYPE: nucleic acid

Ala Ala Gln Met Leu Glu Val Gln Asn Glu Ala

- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: mRNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

•						
AUGACUACUG	AUACCACUGU	UUUCGUUUCU	GGCGCAACCG	GUUUCAUUGC	UCUACACAUU	60
AUGAACGAUC	UGUUGAAAGC	UGGCUAUACA	GUCAUCGGCU	CAGGUAGAUC	UCAAGAAAAA	120
AAUGAUGGCU	UGCUCAAAAA	AUUUAAUAAC	AAUCCCAAAC	UAUCGAUGGA	AAUUGUGGAA	180
GAUAUUGCUG	CUCCAAACGC	CUUUGAUGAA	GUUUUCAAAA	AACAUGGUAA	GGAAAUUAAG	240
AUUGUGCUAC	ACACUGCCUC	CCCAUUCCAU	UUUGAAACUA	CCAAUUUUGA	AAAGGAUUUA	300
CUAACCCCUG	CAGUGAACGG	UACAAAAUCU	AUCUUGGAAG	CGAUUAAAAA	AUAUGCUGCA	360
GACACUGUUG	AAAAAGUUAU	UGUUACUUCG	UCUACUGCUG	CUCUGGUGAC	ACCUACAGAC	420
AUGAACAAAG	GAGAUUUGGU	GAUCACGGAG	GAGAGUUGGA	AUAAGGAUAC	AUGGGACAGU	480
UGUCAAGCCA	ACGCCGUUGC	CGCAUAUUGU	GGCUCGAAAA	AGUUUGCUGA	AAAAACUGCU	540
UGGGAAUUUC	UUAAAGAAAA	CAAGUCUAGU	GUCAAAUUCA	CACUAUCCAC	UAUCAAUCCG	600
GGAUUCGUUU	UUGGUCCUCA	AAUGUUUGCA	GAUUCGCUAA	AACAUGGCAU	AAAUACCUCC	660
UCAGGGAUCG	UAUCUGAGUU	AAUUCAUUCC	AAGGUAGGUG	GAGAAUUUUA	UAAUUACUGU	720
GGCCCAUUUA	UUGACGUGCG	UGACGUUUCU	AAAGCCCACC	UAGUUGCAAU	UGAAAAACCA	780
GAAUGUACCG	GCCAAAGAUU	AGUAUUGAGU	GAAGGUUUAU	UCUGCUGUCA	AGAAAUCGUU	840
GACAUCUUGA	ACGAGGAAUU	CCCUCAAUUA	AAGGGCAAGA	UAGCUACAGG	UGAACCUGCG	900
ACCGGUCCAA	GCUUUUUAGA	AAAAAACUCU	UGCAAGUUUG	ACAAUUCUAA	GACAAAAAA	960
CUACUGGGAU	UCCAGUUUUA	CAAUUUAAAG	GAUUGCAUAG	UUGACACCGC	GGCGCAAAUG	1020
UUAGAAGUUC	AAAAUGAAGC	С				1041

## (2) INFORMATION FOR SEQ ID NO:13:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 1044 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO

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(ix) FE	ATURE:
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- (A) NAME/KEY: CDS
- (B) LOCATION: 1..1044
- (D) OTHER INFORMATION: S. cerevisiae YGL039W

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

()(_,	,	202									
							GTT Val 10				48
							TTA Leu				96
							AAT Asn				144
							GAG Glu				192
							CAA Gln				240
							GTT Val 90				288
							GTG Val				336
							GAC Asp				384
							TCT Ser				432
Ser					Glu		TGG Trp				480
				Ala			TAC Tyr 170				528
			Ala				GAG Glu				576

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AAA Lys	TTT Phe	ACG Thr 195	CTA Leu	TCA Ser	ACC Thr	ATC Ile	AAC Asn 200	CCA Pro	GGA Gly	TTT Phe	GTT Val	TTT Phe 205	GGC Gly	CCT Pro	CAG Gln	624
CTA Leu	TTT Phe 210	GCC Ala	GAC Asp	TCT Ser	CTT Leu	AGA Arg 215	AAT Asn	GGA Gly	ATA Ile	AAT Asn	AGC Ser 220	TCT Ser	TCA Ser	GCC Ala	ATT Ile	672
ATT Ile 225	GCC Ala	AAT Asn	TTG Leu	GTT Val	AGT Ser 230	TAT Tyr	AAA Lys	TTA Leu	GGC	GAC Asp 235	AAT Asn	TTT Phe	TAT	AAT Asn	TAC Tyr 240	720
AGT Ser	GGT Gly	CCT Pro	TTT Phe	ATT Ile 245	GAC Asp	GTT Val	CGC Arg	GAT Asp	GTT Val 250	TCA Ser	AAA Lys	GCT Ala	CAT His	TTA Leu 255	CTT Leu	768
GCA Ala	TTT Phe	GAG Glu	AAA Lys 260	CCC Pro	GAA Glu	TGC Cys	GCT Ala	GGC Gly 265	CAA Gln	AGA Arg	CTA Leu	TTC Phe	TTA Leu 270	TGT Cys	GAA Glu	816
GAT Asp	ATG Met	TTT Phe 275	TGC Cys	TCT Ser	CAA Gln	GAA Glu	GCG Ala 280	CTG Leu	GAT Asp	ATC Ile	TTG Leu	AAT Asn 285	GAG Glu	GAA Glu	TTT Phe	864
CCA Pro	CAG Gln 290	TTA Leu	AAA Lys	GGC Gly	AAG Lys	ATA Ile 295	GCA Ala	ACT Thr	GGC Gly	GAA Glu	CCT Pro 300	GGT Gly	AGC Ser	GGC Gly	TCA Ser	912
ACC Thr 305	TTT Phe	TTG Leu	ACA Thr	AAA Lys	AAC Asn 310	TGC Cys	TGC Cys	AAG Lys	TGC Cys	GAC Asp 315	AAC Asn	CGC Arg	AAA Lys	ACC Thr	AAA Lys 320	960
AAT Asn	TTA Leu	TTA Leu	GGA Gly	TTC Phe 325	Gln	TTT	AAT Asn	AAG Lys	TTC Phe 330	AGA Arg	GAT Asp	TGC Cys	ATT Ile	GTC Val 335	GAT Asp	1008
ACT Thr	GCC Ala	TCG Ser	CAA Gln 340	Leu	CTA Leu	GAA Glu	GTT Val	CAA Gln 345	AGT Ser	AAA Lys	AGC Ser					1044

## (2) INFORMATION FOR SEQ ID NO:14:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 348 amino acids
  - (B) TYPE: amino acid
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

Met Thr Thr Glu Lys Thr Val Val Phe Val Ser Gly Ala Thr Gly Phe
1 5 10 15

- Ile Ala Leu His Val Val Asp Asp Leu Leu Lys Thr Gly Tyr Lys Val 20 25 30
- Ile Gly Ser Gly Arg Ser Gln Glu Lys Asn Asp Gly Leu Leu Lys Lys

  45
- Phe Lys Ser Asn Pro Asn Leu Ser Met Glu Ile Val Glu Asp Ile Ala
  50 55 60
- Ala Pro Asn Ala Phe Asp Lys Val Phe Gln Lys His Gly Lys Glu Ile 65 70 75 80
- Lys Val Val Leu His Ile Ala Ser Pro Val His Phe Asn Thr Thr Asp 85 90 95
- Phe Glu Lys Asp Leu Leu Ile Pro Ala Val Asn Gly Thr Lys Ser Ile 100 105 110
- Leu Glu Ala Ile Lys Asn Tyr Ala Ala Asp Thr Val Glu Lys Val Val 115 120 125
- Ile Thr Ser Ser Val Ala Ala Leu Ala Ser Pro Gly Asp Met Lys Asp 130 135 140
- Thr Ser Phe Val Val Asn Glu Glu Ser Trp Asn Lys Asp Thr Trp Glu 145 150 155 160
- Ser Cys Gln Ala Asn Ala Val Ser Ala Tyr Cys Gly Ser Lys Lys Phe 165 170 175
- Ala Glu Lys Thr Ala Trp Asp Phe Leu Glu Glu Asn Gln Ser Ser Ile 180 185 190
- Lys Phe Thr Leu Ser Thr Ile Asn Pro Gly Phe Val Phe Gly Pro Gln 195 200 205
- Leu Phe Ala Asp Ser Leu Arg Asn Gly Ile Asn Ser Ser Ser Ala Ile 210 215 220
- Ile Ala Asn Leu Val Ser Tyr Lys Leu Gly Asp Asn Phe Tyr Asn Tyr 225 230 235 240
- Ser Gly Pro Phe Ile Asp Val Arg Asp Val Ser Lys Ala His Leu Leu 245 250 255
- Ala Phe Glu Lys Pro Glu Cys Ala Gly Gln Arg Leu Phe Leu Cys Glu 260 265 270
- Asp Met Phe Cys Ser Gln Glu Ala Leu Asp Ile Leu Asn Glu Glu Phe 275 280 285
- Pro Gln Leu Lys Gly Lys Ile Ala Thr Gly Glu Pro Gly Ser Gly Ser 290 295 300
- Thr Phe Leu Thr Lys Asn Cys Cys Lys Cys Asp Asn Arg Lys Thr Lys

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305 310 315 320

Asn Leu Leu Gly Phe Gln Phe Asn Lys Phe Arg Asp Cys Ile Val Asp 325 330 335

Thr Ala Ser Gln Leu Leu Glu Val Gln Ser Lys Ser 340 345

#### (2) INFORMATION FOR SEQ ID NO:15:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 1044 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: mRNA
- (iii) HYPOTHETICAL: NO
  - (iv) ANTI-SENSE: NO

#### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:

AUGACUACUG AAAAAACCGU UGUUUUUGUU UCUGGUGCUA CUGGUUUCAU UGCUCUACAC 60 GUAGUGGACG AUUUAUUAAA AACUGGUUAC AAGGUCAUCG GUUCGGGUAG GUCCCAAGAA 120 AAGAAUGAUG GAUUGCUGAA AAAAUUUAAG AGCAAUCCCA ACCUUUCAAU GGAGAUUGUC 180 GAAGACAUUG CUGCUCCAAA CGCUUUUGAC AAAGUUUUUC AAAAGCACGG CAAAGAGAUC 240 AAGGUUGUCU UGCACAUAGC UUCUCCGGUU CACUUCAACA CCACUGAUUU CGAAAAGGAU 300 CUGCUAAUUC CUGCUGUGAA UGGUACCAAG UCCAUUCUAG AAGCAAUCAA AAAUUAUGCC 360 GCAGACACAG UCGAAAAAGU CGUUAUUACU UCUUCUGUUG CUGCCCUUGC AUCUCCCGGA 420 GAUAUGAAGG ACACUAGUUU CGUUGUCAAU GAGGAAAGUU GGAACAAAGA UACUUGGGAA 480 AGUUGUCAAG CUAACGCGGU UUCCGCAUAC UGUGGUUCCA AGAAAUUUGC UGAAAAAACU 540 GCUUGGGAUU UUCUCGAGGA AAACCAAUCA AGCAUCAAAU UUACGCUAUC AACCAUCAAC 600 CCAGGAUUUG UUUUUGGCCC UCAGCUAUUU GCCGACUCUC UUAGAAAUGG AAUAAAUAGC 660 UCUUCAGCCA UUAUUGCCAA UUUGGUUAGU UAUAAAUUAG GCGACAAUUU UUAUAAUUAC 720 AGUGGUCCUU UUAUUGACGU UCGCGAUGUU UCAAAAGCUC AUUUACUUGC AUUUGAGAAA 780 CCCGAAUGCG CUGGCCAAAG ACUAUUCUUA UGUGAAGAUA UGUUUUGCUC UCAAGAAGCG 840

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CUGGAUAUCU	UGAAUGAGGA	AUUUCCACAG	UUAAAAGGCA	AGAUAGCAAC	UGGCGAACCU	900
GGUAGCGGCU	CAACCUUUUU	GACAAAAAAC	UGCUGCAAGU	GCGACAACCG	CAAAACCAAA	960
AAUUUAUUAG	GAUUCCAAUU	UAAUAAGUUC	AGAGAUUGCA	UUGUCGAUAC	UGCCUCGCAA	1020
UUACUAGAAG	UUCAAAGUAA	AAGC				1044

# INTERNATIONAL SEARCH REPORT

International application No. PCT/US98/23419

A. CLASSIFICATION OF SUBJECT MATTER  IPC(6) :C12P 21/06; C12N 9/02, 1/20, 15/00; C07H 21/04; US CL :435/69.1, 189, 252.3, 320.1; 536/23.2, 23.7; 530/350  According to International Patent Classification (IPC) or to both	).					
B. FIELDS SEARCHED						
Minimum documentation searched (classification system follower	d by classification symbols)					
U.S. : 435/69.1, 189, 252.3, 320.1; 536/23.2, 23.7; 530/350.						
Documentation searched other than minimum documentation to the	e extent that such documents are included in the fields searched					
Electronic data base consulted during the international search (na	ame of data base and, where practicable, scatch what allege					
Please See Extra Sheet.						
C. DOCUMENTS CONSIDERED TO BE RELEVANT						
Category* Citation of document, with indication, where ap	propriate, of the relevant passages Relevant to claim No.					
A JORNVALL et al. Short-Chain Dehyd Biochemistry. 09 May 1995. Vol 34, I the entire article.	rogenases/Reductases (SDR). 1-21 No. 18, pages 6003-6013, see					
Further documents are listed in the continuation of Box C						
Special categories of cited documents:  A document defining the general state of the art which is not considered to be of particular relevance.	date and not in conflict with the application but cited to understand the principle or theory underlying the invention					
*E* earlier document published on or after the international filing date	<ul> <li>'X' document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step</li> </ul>					
*L* document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	when the document is taken alone  'Y' document of particular relevance; the claimed invention cannot be					
*O* document referring to an oral disclosure, use, exhibition or other means	considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art					
*P* document published prior to the international filing date but later than the priority date claimed	"A." document member of the same patent family					
Date of the actual completion of the international search	Date of mailing of the international search report 03 FEB 1999					
29 DECEMBER 1998	·					
Name and mailing address of the ISA/US Commissioner of Patents and Trademarks Box PCT Washington, D.C. 20231 Facsimile No. (703) 305-3230	TEKCHAND SAIDHA  Telephone No. (703) 308-0196					

# INTERNATIONAL SEARCH REPORT

International application No. PCT/US98/23419

	B. FIELDS SEARCHED  Electronic data bases consulted (Name of data base and where practicable terms used):				
STN Files: Medline, Caplus, Wpids, Biosis, Biotechds, Scisearch. Search terms included: ketoreductase, Zygosaccharomyces, yeast in combinations as well as authurs names search. APS search.					
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I					